



DOI: <https://doi.org/10.23859/estr-240410>

EDN: <https://elibrary.ru/dvxkks>

UDC 581.19+581.4+632.12+633.161

### Article

# The effect of manganese and cadmium on morphological and biochemical parameters of barley seedlings

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**Abstract.** The influence of various concentrations of  $Mn^{2+}$  (100–800 mg/l) and  $Cd^{2+}$  (10–80 mg/l) on spring barley *Hordeum vulgare* L., variety ‘Rodnik Prikamya’ was studied in the roll culture under laboratory conditions. Seed germination, seedling biomass, and the content of free and bound polyphenols in plant biomass were estimated. The effect of low-dose manganese (100 mg/l) turned out to be stimulating: seed germination and biomass of barley seedlings increased by 35 and 51% compared to the control. Elevated concentrations of  $Mn^{2+}$  (200–800 mg/l) suppressed seed germination and biomass (by 55–75% and 60–79%). Cadmium (10–80 mg/l) made solely inhibitory impact: seed germination decreased by 20–70% and seedling biomass by 23–78%. Biochemical responses of plants to stress caused by these elements differed. For instance, application of  $Mn^{2+}$  increased the proportion of bound polyphenols in the total polyphenol of seedlings by 8, 12, 7, and 8% compared to the control. Cadmium stress was compensated by polyphenols only at its lowest tested concentration (20 mg/L); the proportion of bound polyphenols increased by 3.3%, as compared to the control. In case with the highest concentrations of the studied metal ions (500–800 mg/L for  $Mn^{2+}$  and 50–80 mg/L for  $Cd^{2+}$ ), the content of polyphenols dropped to control values thereby indicating the depletion of adaptive reserves of plants.

**Keywords:** phytotesting, *Hordeum vulgare*, biomass, germination, polyphenols, toxicity

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**To cite this article:** Tovstik, E.V., Olkova, A.S., 2025. The effect of manganese and cadmium on morphological and biochemical parameters of barley seedlings. *Ecosystem Transformation* 8 (3), 198–208. <https://doi.org/10.23859/estr-240410>

Received: 10.04.2024

Accepted: 02.08.2024

Published online: 29.08.2025

DOI: <https://doi.org/10.23859/estr-240410>EDN: <https://elibrary.ru/dvxkks>

УДК 581.19+581.4+632.12+633.161

**Научная статья****Влияние марганца и кадмия на морфофизиологические и биохимические показатели проростков ячменя**Е.В. Товстик<sup>ORCID</sup>, А.С. Олькова\*<sup>ORCID</sup>*Вятский государственный университет, Россия, 610000, г. Киров, ул. Московская, д. 36**\*morgan-abend@mail.ru*

**Аннотация.** В лабораторном опыте исследовали действие различных концентраций  $Mn^{2+}$  (100–800 мг/л) и  $Cd^{2+}$  (10–80 мг/л) на растения ярового ячменя *Hordeum vulgare* L., сорт 'Родник Прикамья'. Опыт проводили в рулонной культуре. Оценивали всхожесть семян, биомассу проростков, содержание свободных и связанных полифенолов в биомассе растений. Эффект марганца в низкой концентрации (100 мг/л) был стимулирующим: всхожесть семян и биомасса проростков ячменя увеличилась на 35 и 51% по сравнению с контролем. Повышенная концентрация  $Mn^{2+}$  (200–800 мг/л) ингибировала всхожесть семян и биомассу проростков ячменя (на 55–75% и 60–79%). Действие  $Cd^{2+}$  в пределах 10–80 мг/л было только угнетающим: всхожесть семян снизилась на 20–70%, биомасса проростков – на 23–78%. Биохимические реакции растений на стресс, вызванный марганцем и кадмием, различались. Марганец вызывал увеличение доли связанных полифенолов в общем полифенольном профиле проростков на 8, 12, 7 и 8% по сравнению с контролем. Кадмиевый стресс был компенсирован полифенолами только в наименьшей тестируемой концентрации (20 мг/л): доля связанных полифенолов возросла на 3.3% к контролю. При наиболее высоких концентрациях ионов исследуемых металлов (500–800 мг/л  $Mn^{2+}$  и 50–80 мг/л  $Cd^{2+}$ ) происходило снижение содержания полифенолов до контрольных значений, что свидетельствует об истощении адаптационных резервов растений.

**Ключевые слова:** фитотестирование, *Hordeum vulgare*, биомасса, всхожесть, полифенолы, токсичность

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**Для цитирования:** Товстик, Е.В., Олькова, А.С., 2025. Влияние марганца и кадмия на морфофизиологические и биохимические показатели проростков ячменя. *Трансформация экосистем* 8 (3), 198–208. <https://doi.org/10.23859/estr-240410>

Поступила в редакцию: 10.04.2024

Принята к печати: 02.08.2024

Опубликована онлайн: 29.08.2025

## Introduction

In laboratory biotesting, plants are often used as test organisms due to their high sensitivity to a wide range of environmental conditions (Agrawal and Agrawal, 1999; Qaderi et al., 2023). Studies suggest that plants quickly respond to changes in lighting, temperature, humidity, carbon dioxide concentration and other parameters (Driesen et al., 2020). In plant cultivation, the assessment of characteristics of seed germination and further growth of seedlings is an integral part of diagnostics of plant resistance to various stress factors (Alekseychuk and Laman, 2005).

Both, in national and international practices, numerous certified phytotesting methods are based on accounting the alterations of major morphophysiological parameters of plants. Among them, seed germination and root length of plants are most often assessed (Terekhova et al., 2016). Suppression of seed germination and root length reduction were recorded in response to oil and salt pollution (Arzamazova et al., 2020), while inhibition of root and shoot elongation – to antibiotics present in soil (Kosmacheva, 2020). Toxicity and dose-dependent effect of fungicides (Saneeva et al., 2022), heavy metals (Alyabysheva, 2023), contaminated sludge sediments (Bostubaeva and Nauanova, 2022), and other toxicants (Koval and Ogorodnikova, 2023) were determined from a set of germination indicators, i.e. the seed germination energy, root and shoot length of plants. Therefore, the use of plants in the environmental monitoring of technogenically disturbed soils (Krasnoperova, 2015), urbanozems (Zykova et al., 2017), natural ecosystems and agrocenoses (Morozova et al., 2020) has been scientifically substantiated.

In biotesting, the employment of morphophysiological reactions of plants to stress factors as test functions is considered to be a simple and reliable approach. However, various stressors can be responsible for a reduced germination, slower growth of roots and seedlings, their external damage and death of plants at the beginning of ontogenesis. It is worth noting that biochemical responses are the first stage of plant adaptation to the changing environment, preceding the morphophysiological transformations. When stress is low, biochemical alterations may occur without external deviations in plant development. In some cases, biochemical indicators form specific combinations corresponding to a certain impact or its level. This is the scientific basis for biomarking. Radicals (reactive oxygen species), ions ( $\text{Na}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{NO}_3^-$ ,  $\text{NH}_4^+$ ,  $\text{H}^+$ ), inorganic ( $\text{H}_2\text{O}$ ,  $\text{CO}_2$ ) and organic molecules (phytohormones, volatile compounds, pigments, RNA molecules) can serve as plant biomarkers (Tan et al., 2023).

Polyphenols are among the low-molecular secondary metabolites that exert a protective effect in response to biotic and abiotic stresses. The spectrum of polyphenolic compounds in plants is quite wide. In plant tissues, they exist both in the form of glycosides (bound fraction) and aglycones (free fraction).

The role of polyphenols in plant resistance has not been fully understood. On the one hand, it is associated with the species- and variety specificity of plants. At the same time, non-specific hormetic activation of various protective systems by low concentrations of toxicants is insufficiently studied (Erofeeva, 2022). Of practical interest is also a comparison of classical morphophysiological responses of plants to chemical stress and biochemical changes in plant test organisms.

Despite the accumulated data on characteristics of individual heavy metals and their impact on living organisms, identification of differences and similarities in the behavior of these elements still remains an urgent task in solving the problems of environmental pollution.

This study is aimed at the comparison of morphophysiological and biochemical reactions of spring barley seedlings *Hordeum vulgare* L. to various concentrations of manganese and cadmium in order to supplement the information about the mechanisms of plants' protection from chemical effects.

## Materials and methods

### **Model experiment**

In laboratory experiments, we studied the influence of various concentrations of manganese and cadmium ions on spring barley seedlings *Hordeum vulgare* L. For testing, barley seeds of the 'Rodnik Prikamya' variety (originator: FGBNU FANTS Severo-Vostok) were used. Laboratory seed germination made up 92%<sup>1</sup>.

Phytotesting was implemented according to the method described in GOST 12038-84 with some modifications. Seeds were germinated in the roll culture. Before testing, barley seeds were soaked

<sup>1</sup> GOST 12038-84 Seeds of agricultural crops. Methods for determining germination.

in a distilled water for 16 hours. Then, swollen seeds (35 pcs.) were laid out on sheets of filter paper (15×50 cm), preheated in a dry-heat oven for 2 hours at a temperature of 105 °C. Seeds were oriented with the embryos down (2–3 cm from the upper edge) and covered with a similarly prepared filter paper of 5×50 cm size. Paper strips with seeds were loosely rolled and placed vertically in 500 ml glass beakers filled with salt solutions of manganese ( $\text{MnSO}_4 \cdot 5 \text{H}_2\text{O}$ ) and cadmium ( $\text{CdSO}_4 \cdot 2.7 \text{H}_2\text{O}$ ) of various concentrations (Table 1).

Cadmium was chosen for experiments due to its toxicity and hazard for human health (Wang et al., 2021), while manganese because of its prevalence and high content in soils (from 221 to 1428 mg/kg) (Simonova et al., 2019). Modeling of the increased manganese concentration was explained by its lower hazard class (class 3) compared to cadmium (class 1)<sup>2</sup>.

A distilled water served as a laboratory control. Seed germination and subsequent plant cultivation were carried out at room temperature of  $20 \pm 2$  °C with a photoperiod of 16 hours of light / 8 hours of darkness. The 10-day experiment was repeated three times.

### ***Evaluated indicators***

Following the exposure, we removed seedlings from rolls, counted normally germinated seeds and estimated raw biomass of seedlings.

After drying and grinding the sprouts, a combined sample of plant material was prepared, and polyphenols were extracted. The total content of polyphenols was determined in alkaline (2N NaOH solution,  $t = 80$  °C,  $\tau = 2$  h) and free polyphenols in aqueous-alcoholic extracts (70%  $\text{C}_2\text{H}_5\text{OH}$  solution,  $t = 5$  °C,  $\tau = 16$  h). The substrate/extractant ratio was 1 : 100. The polyphenol content was defined spectrophotometrically<sup>3</sup>. Gallic acid was employed as a standard. The mass fraction of bound (glycoside) polyphenol forms was found from the difference between the total content and free fraction of polyphenols.

### ***Reliability and mathematical treatment of results***

Each experimental variant was performed in triplicate with the result represented as the mean value and its standard deviation.

Marked on the graphs with Latin letters, significance of differences between the data sets was proved by the one-way analysis of variance (ANOVA) method. The significance level made up 0.05.

Pearson correlation coefficients ( $r$ ) were calculated using Microsoft Excel 2007.

## **Results**

### ***Seed germination and biomass of barley seedlings under chemical stress***

The negative effect of  $\text{Mn}^{2+}$  and  $\text{Cd}^{2+}$  on barley plants was determined from the reduced (compared to the control) seed germination and seedling biomass (Figs. 1A, B). Note that with the increase in ions concentration, the studied parameters in solutions decreased.

In experiments with the lowest concentration of  $\text{Mn}^{2+}$  (100 mg/l), we observed a stimulating effect expressed in the growing seed germination and seedling biomass (by 35 and 51%, respectively). At higher concentrations (200–800 mg/l), manganese, on the contrary, negatively affected the morphophysiological indices of barley growth, i.e. seed germination dropped by 25–75% and seedling biomass by 21.4–79%.

The influence of  $\text{Cd}^{2+}$  and  $\text{Mn}^{2+}$  on test organisms was similar. However, in the experiment with cadmium, all the studied concentrations made an inhibitory effect on seed germination (by 20–70%) and seedling biomass (by 23–78%). It should be noted that the obtained morphophysiological parameters significantly differed from the control values in all experiments, except for that with the lowest tested concentration of  $\text{Cd}^{2+}$  (10 mg/l).

The conducted series of experiments suggests that the selected dose ranges for  $\text{Mn}^{2+}$  and  $\text{Cd}^{2+}$  are efficient, i.e. they do make an effect on test organisms. The use of the same bioassays has made it possible to find out the way of changing the phenolic status of a phytoobject.

<sup>2</sup> GOST 17.4.1.02-83. Environmental protection, classification of chemicals for pollution control.

<sup>3</sup> GOST R 55488-2013. Propolis. Method for determining polyphenols.

**Table 1.** Experimental variants. Dash means the absence of a standard.

Ion	Estimated concentration, mg/l				MPC / APC in soil (total form), mg/kg <sup>4</sup>
Mn(II)	100	200	500	800	1500 / –
Cd(II)	10	20	50	80	– / 0.5–2.0

### ***Phenolic status of barley under chemical stress***

It is known that inhibition of barley seedling growth influenced by heavy metals largely occurs due to non-specific oxidative stress (Juknys et al., 2012). In this regard, among the studied parameters, polyphenols, as markers of chemical stress, are of particular interest.

Manganese and cadmium in the selected concentration ranges had a similar effect on the polyphenol profile of barley seedlings (Figs. 2A, B).

In experiments with the lowest concentrations (100 mg/l for Mn<sup>2+</sup> and 10 mg/l for Cd<sup>2+</sup>), no significant differences in the polyphenol content in barley biomass were recorded. With a 2-fold increase in the concentration (relative to the lowest one), the studied parameter significantly increased, as compared to the control (by 4 and 2.2 mg/g, respectively).

At higher concentrations of Mn<sup>2+</sup> in the medium, the content of total polyphenolic compounds in biomass dropped to the control level. In the variant with maximum Cd<sup>2+</sup> concentrations (80 mg/l), a tendency towards a decrease (compared to the control) in the study parameter was noted.

To clarify the mechanism of biochemical adaptation of plants to chemical stress, the proportional ratio of free and bound polyphenols in their total pool was estimated (Figs. 3A, B).

In the experiment with Mn<sup>2+</sup>, as its concentration increased, the proportion of free polyphenols in the total content significantly decreased, whereas the share of bound ones in seedlings, on the contrary, increased (by 8, 12, 7, 8% compared to the control).

Bearing in mind that free polyphenols have a higher complex-forming potential than the bound ones (Eghbaliferiz and Iranshahi, 2016), excess manganese could bind to aglycones and increase the proportion of the bound fraction, including the total polyphenols in seedlings (Fig. 2).

Unlike manganese, cadmium did not have a significant effect on the ratio of polyphenol fractions. The exception was the experiment with the lowest concentration of Cd<sup>2+</sup> in the solution (10 mg/l). Here, the proportion of free polyphenols in barley seedlings increased by 3.3% compared to the control.

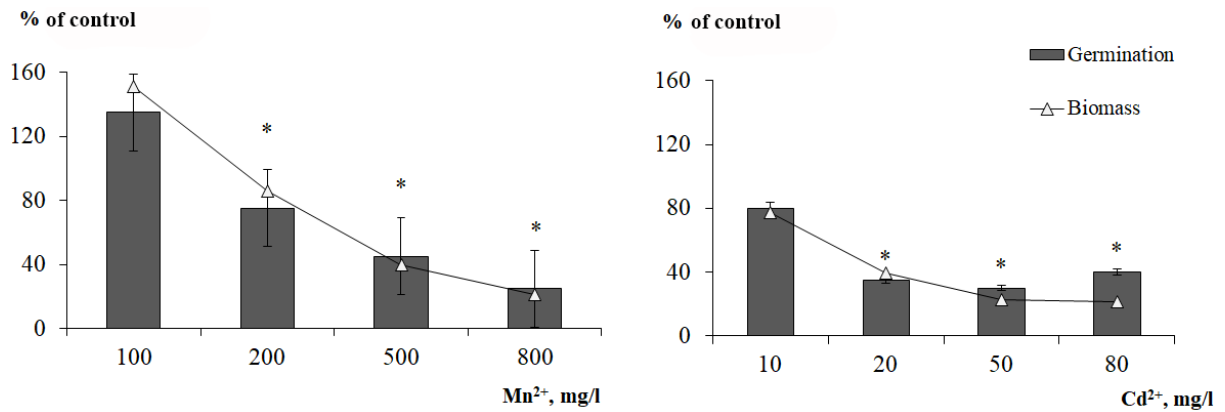
### **Discussion**

The roll culture-based phytotesting in the medium with manganese and cadmium sulfate solutions (100–800 and 10–80 mg/l by metal cations) demonstrates that seed germination and biomass of barley seedlings evidently and consistently change with increasing concentrations of the studied metal ions. In response to Cd<sup>2+</sup> and Mn<sup>2+</sup> effects, biomass ( $r = -0.82$  and  $r = -0.86$ ) and seedling germination ( $r = -0.82$  and  $r = -0.86$ ) significantly decrease.

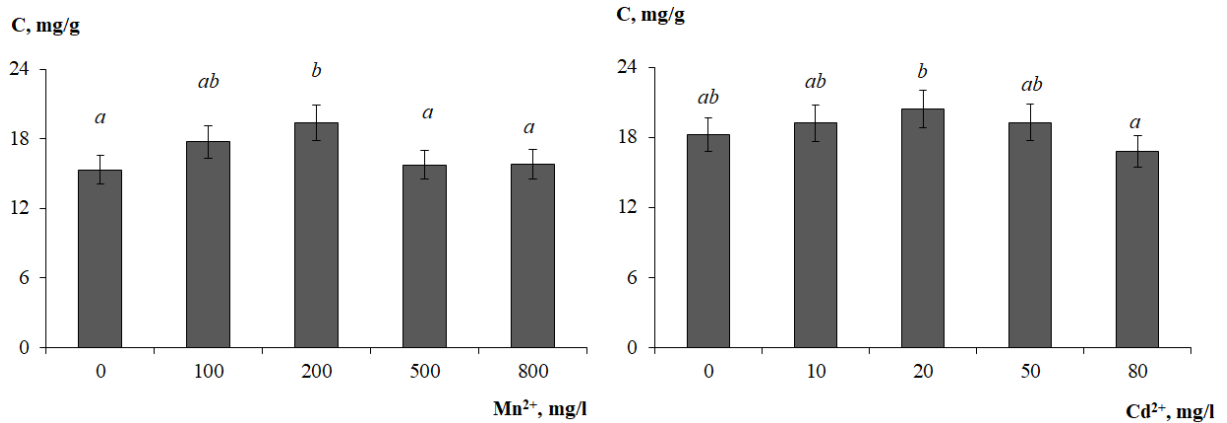
In terms of total polyphenols and its components, the biochemical reaction of barley appears to be smoothed (as compared to the morphophysiological indexes) that is logical since the reaction norm of plant growth characteristics within one species is, of course, wider than that of biochemical indicators ensuring internal homeostasis. However, this fact makes the identified fluctuations in the phenolic status of phytoobjects extremely important. With a consistent elevation of concentrations of Mn<sup>2+</sup> (to 200 mg/l) and Cd<sup>2+</sup> (to 20 mg/l), the total content of polyphenols in seedlings grows. It is due to the property of plants to increase the content of polyphenolic compounds under stress conditions (Tuladhar et al., 2021). The researchers explain this phenomenon by the pronounced antioxidant features of polyphenols, protecting cells from oxidative damage by metals (Chen et al., 2020).

In the culture medium, a subsequent increase in concentrations of Mn<sup>2+</sup> and Cd<sup>2+</sup> (above 200 mg/l and 20 mg/l, respectively) results in the total polyphenols decline in seedlings to the control level.

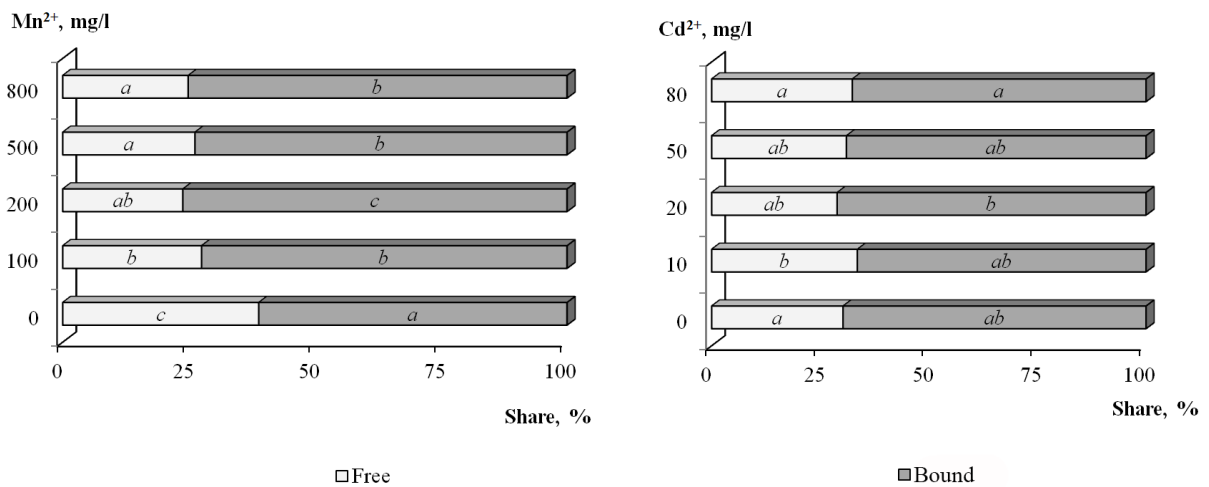
<sup>4</sup> SanPiN 1.2.3685-21. On approval of sanitary rules and regulations "Hygienic standards and requirements for ensuring safety and (or) harmlessness of environmental factors for humans".



**Fig. 1.** Changes in morphophysiological parameters of barley sprout growth influenced by various concentrations of manganese and cadmium. \* – a significant level of differences in parameters compared to the control ( $p < 0/05$ ).



**Fig. 2.** Changes in the total content of polyphenols in barley sprouts influenced by various concentrations of manganese and cadmium. Different letters indicate significant differences ( $p < 0.05$ ).



**Fig. 3.** The ratio of free and bound fractions of polyphenols in barley sprouts influenced by various concentrations of manganese and cadmium. Different letters indicate significant differences ( $p < 0.05$ ).

Apparently, chemical loads (that exceed the adaptive plant reserves) bring to a complex disruption of the antioxidant system and biosynthesis slowdown of polyphenols (Goncharuk and Zagoskina, 2023).

The mechanism of the biochemical response to a chemical stress shows low correlation coefficients with the studied metal concentrations:  $r = -0.023$  ( $p = 0.93$ ) for  $Mn^{2+}$  and  $r = -0.07$  ( $p = 0.82$ ) for  $Cd^{2+}$ .

The distribution of free and bound forms of polyphenols in their total pool differed, depending on active ions. As a biogenic element, manganese was neutralized by barley through the complexation with polyphenols resulting in the increase of bound polyphenols compared to free ones. Such a pattern was not established in experiments with cadmium. This fact and the tendency towards the falling content of total polyphenols in plants (Fig. 2) against the background of significant inhibition of morphophysiological indicators (Fig. 1) is evidence of a plant inability to neutralize the toxic effect of cadmium because of its binding with polyphenols. Thus, as a xenobiotic, cadmium disrupts the adaptive capabilities of the organism.

It is common knowledge that induction of biosynthesis of non-enzymatic antioxidants may depend on the type of metal and its concentration. Previously, in experiments with wheat seedlings, anthocyanin production rise was detected under moderate cadmium ( $25 \mu\text{mol CdCl}_2$ ) – induced stress, whereas suppression of their antioxidant properties and drop in protective effect under severe one ( $50 \mu\text{mol CdCl}_2$ ) (Shoeva and Khlestkina, 2018).

## Conclusion

The conventional group of chemical elements classified as heavy metals includes both essential elements necessary for living organisms (Mn, Cu, Zn) and highly toxic ones (Cd, Pb, Hg). The significance of the latter for biotas is not fully understood. In this study, an attempt was made to compare the effect of needed for living organism manganese and cadmium, being at the same time potentially toxic even in low concentrations. When exposed to  $Mn^{2+}$  (100–800 mg/l) and  $Cd^{2+}$  (10–80 mg/l) solutions, biomass of barley seedlings decreased and polyphenol profile changed.

The sensitivity of barley seedlings to increasing concentrations of  $Mn^{2+}$  and  $Cd^{2+}$  was revealed from transformations of morphophysiological indexes. For instance, the correlation coefficients of "germination-concentration" and "biomass-concentration" parameters made up  $r = -0.90$  and  $r = -0.55$  for manganese,  $r = -0.91$  and  $r = -0.81$  for cadmium, respectively. Therefore, an excessive essential element (manganese) in plants causes the reactions similar to the effects of a highly toxic element (cadmium).

A difference in biochemical reactions of plants to stress induced by  $Mn^{2+}$  and  $Cd^{2+}$  is shown. In barley seedlings, manganese increases the proportion of bound polyphenols in the total polyphenol profile (by 8, 12, 7 and 8% for successively increasing concentrations). This suggests the antioxidant system response to manganese stress and worsening of its performance with growing chemical loads. Cadmium provides the increased proportion of bound polyphenols only at the lowest tested concentration (by 3.3% compared to the control). At maximum concentrations of  $Cd^{2+}$  (500–800 mg/l and 50–80 mg/l, respectively), the content of polyphenols drops to control values thereby indicating the depletion of adaptive reserves of plants.

Thus, the mechanisms of the toxic effect of elevated concentrations of  $Mn^{2+}$  and  $Cd^{2+}$  were revealed due to morphophysiological and biochemical reactions of plants.

## References

- Agrawal, M., Agrawal, S.B., 1999. Effects of air pollution on plant diversity. In: S.B. Agrawal, M. Agrawal (eds.), *Environmental pollution and plant responses*. Lewis, New York, USA, 137–152.
- Alekseichuk, G.N., Laman, N.A., 2005. Fiziologicheskoe kachestvo semyan sel'skohozyaistvennykh kul'tur i metody ego otsenki [Physiological quality of seeds of agricultural crops and methods of its assessment]. Pravo i ekonomika, Minsk, Belarus', 48 p. (In Russian).
- Alyabysheva, E.A., 2023. Vliyaniye solei svintsa na morfofiziologicheskie pokazateli donnika belogo (*Melilotus albus* Medik.) [The influence of lead salts on the morphophysiological parameters of white sweet clover (*Melilotus albus* Medik.)]. *Vestnik Mariiskogo gosudarstvennogo universiteta. Seriya "Sel'skohozyajstvennyye nauki. Ekonomicheskie nauki"* [Bulletin of the Mari State University.

*Series "Agricultural sciences. Economic sciences"* **9** (2), 131–138. (In Russian). <https://doi.org/10.30914/2411-9687-2023-9-2-131-138>

Arzamazova, A.V., Kinzhaev, R.R., Beletskaya, D.V., 2020. Otsenka toksichnosti neftezagryaznennykh pochv po pokazatelyam vskhozhesti semyan i dliny kornei prorstkov pshenitsy (*Triticum aestivum* L.). [Evaluation of toxicity of oil-contaminated soils based on seed germination and root length of wheat seedlings (*Triticum aestivum* L.)]. *Problemy agrokhimii i ekologii [Problems of Agrochemistry and Ecology]* **4**, 51–55. (In Russian). <https://doi.org/10.26178/AE.2020.51.76.007>

Bostubaeva, M.B., Nauanova, A.P., 2022. Otsenka fitotoksichnosti i rostostimuliruiushchikh svoystv razlichnykh kontsentratsii ilovykh osadkov stochnykh vod na prorstki l'na maslichnogo [Evaluation of phytotoxicity and growth-promoting properties of different concentrations of wastewater sludge on oil flax sprouts]. *Vestnik nauki Kazakhskogo agrotekhnicheskogo universiteta im. S. Seifullina [Herald of Science of S. Seifullin Kazakh Agro Technical Research University]* **2** (113), 170–178. (In Russian). [https://doi.org/10.51452/kazatu.2022.2\(113\).1030](https://doi.org/10.51452/kazatu.2022.2(113).1030)

Chen, Y., Huang, L., Liang, X., Dai, P., Zhang, Y. et al., 2020. Enhancement of polyphenolic metabolism as an adaptive response of lettuce (*Lactuca sativa*) roots to aluminum stress. *Environmental Pollution* **261**, 114230. <https://doi.org/10.1016/j.envpol.2020.114230>

Driesen, E., Van den Ende, W., De Proft, M., Saeys, W., 2020. Influence of environmental factors light, CO<sub>2</sub>, temperature, and relative humidity on stomatal opening and development: a review. *Agronomy* **10** (12), 1975. <https://doi.org/10.3390/agronomy10121975>

Eghbaliferiz, S., Iranshahi, M., 2016. Prooxidant activity of polyphenols, flavonoids, anthocyanins and carotenoids: Updated review of mechanisms and catalyzing metals. *Phytotherapy Research* **30**, 1379–1391.

Erofeeva, E.A., 2022. Environmental hormesis of non-specific and specific adaptive mechanisms in plants. *Science of The Total Environment* **804**, 150059. <https://doi.org/10.1016/j.scitotenv.2021.150059>

Goncharuk, E.A., Zagorskina, N.V., 2023. Heavy metals, their phytotoxicity, and the role of phenolic antioxidants in plant stress responses with focus on cadmium: review. *Molecules* **28**, 3921. <https://doi.org/10.3390/molecules28093921>

Juknys, R., Vitkauskaitė, G., Račaitė, M., Vencloviėnė, J., 2012. The impacts of heavy metals on oxidative stress and growth of spring barley. *Central European Journal of Biology* **7**, 299–306. <https://doi.org/10.2478/s11535-012-0012-9>

Koval, E.V., Ogorodnikova, S.Yu., 2023. Treatment of seeds with cyanobacterial biofilm to increase plant resistance to methylphosphonate pollution. *Ecosystem Transformation* **6** (1), 3–11. <https://doi.org/10.23859/estr-220609>

Kosmacheva, A.G., 2020. Issledovanie toksichnosti dernovo-podzolistoi pochvy, zagryaznennoi antibiotikami razlichnykh grupp, metodom laboratornogo fitotestirovaniya [Study of toxicity of sod-podzolic soil contaminated with antibiotics of various groups using laboratory phytotesting method]. *Sbornik materialov V Mezhdunarodnoi nauchno-prakticheskoi konferentsii "Transgranichnoe sotrudnichestvo v oblasti ekologicheskoi bezopasnosti i okhrany okruzhayushchei sredy" [Collection of materials of the V International scientific and practical conference "Cross-border cooperation in the field of environmental safety and protection"]*. Gomel', Belarus', 290–296. (In Russian).

Krasnoperova, S.A., 2015. Morfologicheskii analiz i rezistentnost' rastenii, rekomenduemykh dlya fitoremediatsii neftezagryaznennykh pochv [Morphological analysis and resistance of plants recommended

for phytoremediation of oil-contaminated soils]. *Sovremennye naukoemkie tekhnologii. Regional'noe prilozhenie [Modern Science-Intensive Technologies. Regional Application]* 4 (44), 184–188. (In Russian).

Morozova, T.S., Shiryaev, A.V., Timofeev, T.A., 2020. Agroekologicheskaya otsenka fitotoksichnosti pochv estestvennykh tsenozov i agrotsenoza [Agroecological assessment of phytotoxicity of soils of natural cenoses and agrocenoses]. *Innovacii v APK: problemy i perspektivy [Innovations In the Agro-Industrial Complex: Problems and Prospects]* 2, 185–190. (In Russian).

Qaderi, M.M., Martel, A.B., Strugnell, C.A., 2023. Environmental factors regulate plant secondary metabolites. *Plants* 12 (3), 447. <https://doi.org/10.3390/plants12030447>

Saneeva, E.A., Zor'kina, O.V., Nefed'eva, E.E., 2022. Issledovanie fitotoksicheskogo deistviya tebukonazola, protiokonazola, fludioksonila i preparatov na ikh osnove na energiyu prorastaniya i rost prorostkov pshenitsy i gorchitsy beloi [Study of phytotoxic effect of tebuconazole, prothioconazole, fludioxonil and preparations based on them on germination energy and growth of wheat and white mustard seedlings]. *Siberian Journal of Life Sciences and Agriculture* 14 (5), 166–186. (In Russian). <https://doi.org/10.12731/2658-6649-2022-14-5-166-186>

Shoeva, O.Y., Khlestkina, E.K., 2018. Anthocyanins participate in the protection of wheat seedlings against cadmium stress. *Cereal Research Communications* 46 (2), 242–252. <https://doi.org/10.1556/0806.45.2017.070>

Simonova, O.A., Lisitsyn, E.M., Tovstik, E.V., 2019. Sravnitel'noe sodержanie margantsa v verkhnikh gorizontakh pochv Kirovskoi oblasti [Comparative content of manganese in the upper soil horizons of the Kirov region]. *Estestvennye i tekhnicheskie nauki [Natural and Technical Sciences]* 10 (136), 127–131. (In Russian). <https://doi.org/10.25633/ETN.2019.10.23>

Tan, Y., Duan, Y., Chi, Q., Wang, R., Yin, Y. et al., 2023. The role of reactive oxygen species in plant response to rRadiation. *International Journal of Molecular Sciences* 24 (4), 3346. <https://doi.org/10.3390/ijms24043346>

Terekhova, V.A., Voronina, L.P., Nikolaeva, O.V., Bardina, T.V., Kalmackaya, O.A. et al., 2016. Primenenie fitotestirovaniya dlya resheniya zadach ekologicheskogo pochvovedeniya [Application of phytotesting to solve problems of ecological soil science]. *Ispol'zovanie i ohrana prirodnih resursov v Rossii [Use and Protection of Natural Resources in Russia]* 3, 37–41. (In Russian).

Tuladhar, P., Sasidharan, S., Saudagar, P., 2021. 17 – Role of phenols and polyphenols in plant defense response to biotic and abiotic stresses. In: Jogaiyah, S. (ed.), *Biocontrol Agents and Secondary Metabolites. Applications and Immunization for Plant Growth and Protection*. Elsevier, Amsterdam, The Netherlands, 419–441. <https://doi.org/10.1016/B978-0-12-822919-4.00017-X>

Wang, M., Chen, Z., Song, W., Hong, D., Huang, L., Li, Y., 2021. A review on cadmium exposure in the population and intervention strategies against toxicity. *Bulletin of Environmental Pollution and Toxicology* 106 (1), 65–74. <https://doi.org/10.1007/s00128-020-03088-1>

Zykova, Yu.N., Skugoreva, S.G., Tovstik, E.V., Ashihmina, T.Ya., 2017. Podkhody k otsenke sostoyaniya gorodskikh pochv metodami biotestirovaniya s ispol'zovaniem organizmov razlichnoi sistemicheskoi prinadlezhnosti i dannykh khimicheskogo analiza [Approaches to assessing the state of urban soils by biotesting methods using organisms of different taxonomic affiliations and chemical analysis data]. *Teoreticheskaya i prikladnaya ekologiya [Theoretical and Applied Ecology]* 3, 38–46. (In Russian). <https://doi.org/10.25750/1995-4301-2017-3-038-046>

## Список литературы

- Алексейчук, Г.Н., Ламан, Н.А., 2005. Физиологическое качество семян сельскохозяйственных культур и методы его оценки. Право и экономика, Минск, Беларусь, 48 с.
- Алябышева, Е.А., 2023. Влияние солей свинца на морфофизиологические показатели донника белого (*Melilotus albus* Medik.). *Вестник Марийского государственного университета. Серия «Сельскохозяйственные науки. Экономические науки»* 9 (2), 131–138. <https://doi.org/10.30914/2411-9687-2023-9-2-131-138>
- Арзамазова, А.В., Кинжаев, Р.Р., Белецкая, Д.В., 2020. Оценка токсичности нефтезагрязненных почв по показателям всхожести семян и длины корней проростков пшеницы (*Triticum aestivum* L.). *Проблемы агрохимии и экологии* 4, 51–55. <https://doi.org/10.26178/AE.2020.51.76.007>
- Бостубаева, М.Б., Науанова, А.П., Nauanova, A.P., 2022. Оценка фитотоксичности и ростостимулирующих свойств различных концентрации иловых осадков сточных вод на проростки льна масличного. *Herald of Science of S. Seifullin Kazakh Agro Technical Research University* 2 (113), 170–178. [https://doi.org/10.51452/kazatu.2022.2\(113\).1030](https://doi.org/10.51452/kazatu.2022.2(113).1030)
- Зыкова, Ю.Н., Скугорева, С.Г., Товстик, Е.В., Ашихмина, Т.Я., 2017. Подходы к оценке состояния городских почв методами биотестирования с использованием организмов различной систематической принадлежности и данных химического анализа. *Теоретическая и прикладная экология* 3, 38–46. <https://doi.org/10.25750/1995-4301-2017-3-038-046>
- Коваль, Е.В., Огородникова, С.Ю., 2023. Обработка семян биопленками цианобактерий для повышения устойчивости растений в условиях химического загрязнения метилфосфонатами. *Трансформация экосистем* 6 (1), 3–11. <https://doi.org/10.23859/estr-220609>
- Космачева, А.Г., 2020. Исследование токсичности дерново-подзолистой почвы, загрязненной антибиотиками различных групп, методом лабораторного фитотестирования. *Сборник материалов V Международной научно-практической конференции "Трансграничное сотрудничество в области экологической безопасности и охраны окружающей среды."* Гомель, Беларусь, 290–296.
- Красноперова, С.А., 2015. Морфологический анализ и резистентность растений, рекомендуемых для фиторемедиации нефтезагрязненных почв. *Современные наукоемкие технологии. Региональное приложение* 4 (44), 184–188.
- Морозова, Т.С., Ширяев, А.В., Тимофеев, Т.А., 2020. Агроэкологическая оценка фитотоксичности почв естественных ценозов и агроценоза. *Инновации в АПК: проблемы и перспективы* 2, 185–190.
- Санеева, Е.А., Зорькина, О.В., Нефедьева, Е.Э., 2022. Исследование фитотоксического действия тебуконазола, протиоконазола, флудиоксонила и препаратов на их основе на энергию прорастания и рост проростков пшеницы и горчицы белой. *Siberian Journal of Life Sciences and Agriculture* 14 (5), 166–186. <https://doi.org/10.12731/2658-6649-2022-14-5-166-186>
- Симонова, О.А., Лисицын, Е.М., Товстик, Е.В., 2019. Сравнительное содержание марганца в верхних горизонтах почв Кировской области. *Естественные и технические науки* 10 (136), 127–131. <https://doi.org/10.25633/ETN.2019.10.23>
- Терехова, В.А., Воронина, Л.П., Николаева, О.В., Бардина, Т.В., Калмацкая, О.А. и др., 2016. Применение фитотестирования для решения задач экологического почвоведения. *Использование и охрана природных ресурсов в России* 3, 37–41.
- Agrawal, M., Agrawal, S.B., 1999. Effects of air pollution on plant diversity. In: S.B. Agrawal, M. Agrawal (eds.), *Environmental pollution and plant responses*. Lewis, New York, USA, 137–152.

- Chen, Y., Huang, L., Liang, X., Dai, P., Zhang, Y. et al., 2020. Enhancement of polyphenolic metabolism as an adaptive response of lettuce (*Lactuca sativa*) roots to aluminum stress. *Environmental Pollution* **261**, 114230. <https://doi.org/10.1016/j.envpol.2020.114230>
- Driesen, E., Van den Ende, W., De Proft, M., Saeys, W., 2020. Influence of environmental factors light, CO<sub>2</sub>, temperature, and relative humidity on stomatal opening and development: a review. *Agronomy* **10** (12), 1975. <https://doi.org/10.3390/agronomy10121975>
- Eghbaliferiz, S., Iranshahi, M., 2016. Prooxidant activity of polyphenols, flavonoids, anthocyanins and carotenoids: Updated review of mechanisms and catalyzing metals. *Phytotherapy Research* **30**, 1379–1391.
- Erofeeva, E.A., 2022. Environmental hormesis of non-specific and specific adaptive mechanisms in plants. *Science of The Total Environment* **804**, 150059. <https://doi.org/10.1016/j.scitotenv.2021.150059>
- Goncharuk, E.A., Zagorskina, N.V., 2023. Heavy metals, their phytotoxicity, and the role of phenolic antioxidants in plant stress responses with focus on cadmium: review. *Molecules* **28**, 3921. <https://doi.org/10.3390/molecules28093921>
- Juknys, R., Vitkauskaitė, G., Račaitė, M., Vencloviėnė, J., 2012. The impacts of heavy metals on oxidative stress and growth of spring barley. *Central European Journal of Biology* **7**, 299–306. <https://doi.org/10.2478/s11535-012-0012-9>
- Qaderi, M.M., Martel, A.B., Strugnell, C.A., 2023. Environmental factors regulate plant secondary metabolites. *Plants* **12** (3), 447. <https://doi.org/10.3390/plants12030447>
- Shoeva, O.Y., Khlestkina, E.K., 2018. Anthocyanins participate in the protection of wheat seedlings against cadmium stress. *Cereal Research Communications* **46** (2), 242–252. <https://doi.org/10.1556/0806.45.2017.070>
- Tan, Y., Duan, Y., Chi, Q., Wang, R., Yin, Y. et al., 2023. The role of reactive oxygen species in plant response to rRadiation. *International Journal of Molecular Sciences* **24** (4), 3346. <https://doi.org/10.3390/ijms24043346>
- Tuladhar, P., Sasidharan, S., Saudagar, P., 2021. 17 – Role of phenols and polyphenols in plant defense response to biotic and abiotic stresses. In: Jogaiah, S. (ed.), *Biocontrol Agents and Secondary Metabolites. Applications and Immunization for Plant Growth and Protection*. Elsevier, Amsterdam, The Netherlands, 419–441. <https://doi.org/10.1016/B978-0-12-822919-4.00017-X>
- Wang, M., Chen, Z., Song, W., Hong, D., Huang, L., Li, Y., 2021. A review on cadmium exposure in the population and intervention strategies against toxicity. *Bulletin of Environmental Pollution and Toxicology* **106** (1), 65–74. <https://doi.org/10.1007/s00128-020-03088-1>